Inflammation and depressive phenotypes: evidence from medical records from over 12 000 patients and brain morphology

Maria Ironside1-2, *, Roee Admon3, *, Stephanie A. Maddox1,2, Malavika Mehta2, Samuel Douglas2, David P. Olson1,2 and Diego A. Pizzagalli1,2

1Department of Psychiatry, Harvard Medical School, Boston, MA 02115, USA; 2McLean Hospital, Belmont, MA 02478, USA and 3Department of Psychology, University of Haifa, Haifa, Israel

Abstract

Background. Preclinical and human studies suggest an association between chronic inflammation and the development of depressive behaviors. This is proposed to occur through downstream effects of inflammatory cytokines on neuroplasticity, neurogenesis and neurotransmitter function, although the neural correlates remain poorly understood in humans.

Methods. In Study 1, structural magnetic resonance imaging and serum inflammatory cytokine data were analyzed from 53 psychiatrically healthy female participants. Correlational analyses were conducted between interleukin-6 (IL-6) and volume in a priori regions implicated in the pathophysiology of major depressive disorder (MDD). In Study 2, medical data [including serum inflammatory acute phase reactants (C-reactive protein)] were analyzed for 12 589 participants. Participants were classified as having (n = 2541) v. not having (n = 10 048) probable lifetime MDD using phenotypes derived using machine-learning approaches. Non-parametric analyses compared inflammation between groups, whereas regression analyses probed whether inflammation predicted probable MDD classification while accounting for other variables.

Results. In Study 1, significant negative correlations emerged between IL-6 and hippocampal, caudate, putamen and amygdalar volume. In Study 2, the MDD group showed a higher probability of elevated inflammation than the non-MDD group. Moreover, elevated inflammation was a significant predictor of probable MDD classification.

Conclusions. Findings indicate that inflammation is cross-sectionally related to reduced volume in brain regions implicated in MDD phenotypes among a sample of psychiatrically healthy women, and is associated with the presence of probable MDD in a large clinical dataset. Future investigations may identify specific inflammatory markers predicting first MDD onset.

Introduction

Major depressive disorder (MDD) is heterogeneous. Accordingly, focusing on intermediate phenotypes (endophenotypes) can be useful to identify more homogenous subgroups of patients sharing common pathophysiology and allow better integration with animal work. Among the most established depressive endophenotypes are anhedonia, stress sensitivity and executive function deficits (Hasler et al., 2004; Goldstein and Klein, 2014; Pizzagalli, 2014). Notably, the increasing availability of large medical databases that can be mined opens new avenues for investigating depressive phenotypes with respect to key pathophysiological variables (Howard et al., 2018).

Growing evidence suggests that chronic, low grade inflammation may induce changes that increase risk for MDD (Miller et al., 2013). In particular, animal studies indicate that pro-inflammatory cytokines [e.g. interleukin-6 (IL-6)] and acute phase reactants [e.g. C-reactive protein (CRP)] have downstream effects, resulting in ‘sickness behaviors’ such as social withdrawal, reduced food consumption and deficits in hippocampal-dependent memory (Dantzer et al., 2008; Iwata et al., 2016; Menard et al., 2017; Wang et al., 2018). Furthermore, elevated concentrations of pro-inflammatory cytokines have been reported in MDD (Dowlati et al., 2010), predicted disease severity (Valkanova et al., 2013) and de novo MDD onset (Pasco et al., 2010) in longitudinal studies, and modulated responses to treatment (Lindqvist et al., 2017). Conversely, and highlighting bidirectional influences, depression has been shown to predict subsequent increases in inflammation (Stewart et al., 2009), which could partly explain the recurrent nature of MDD.

Numerous studies have shown that peripheral inflammation negatively impacts neuronal structure and function, causing neurodegeneration and impeding neuroplasticity (Calabrese et al., 2014) through complex interactions with stress, endocrine and neurotransmitter function (Anisman and Merali, 2003). Studies employing the rodent chronic social defeat stress
paradigm have identified increases in central and peripheral biomarkers of inflammation (Gao et al., 2019; Niraula et al., 2019), whereas ex vivo imaging studies have found increases in volume in nucleus accumbens (NAC) and cingulate cortex, and decreases in ventral tegmental area and amygdala associated with stress response (Anacker et al., 2016). While the mechanisms that mediate volume changes accompanying exposure to stressors remain largely unknown, negative relationships between inflammatory markers and brain volume have emerged in humans, particularly in the hippocampus (Marsland et al., 2008; Frodl et al., 2012; van Velzen et al., 2017) – one of the main sites for neurogenesis. This is interesting given the effects of stress and MDD on hippocampus volume (Belleau et al., 2019) and meta-regression findings highlighting a correlation between number of MDD episodes and decreasing hippocampal volume (Videbech and Ravniilde, 2004; see also Treadway et al., 2015). Bringing this together, studies are beginning to show associations between endogenous inflammation and brain volume in the hippocampus, caudate, amygdala and anterior cingulate cortex (ACC) – regions critically implicated in the pathophysiology of MDD (Savitz et al., 2013; van Velzen et al., 2017).

In humans, substantial evidence for a causal link between the administration of exogenous inflammatory stimuli and the emergence of depressive symptoms can be gleaned from (1) studies involving administration of inflammatory challenges, such as Interferon (IFN)-α treatment for hepatitis C (Bonaccorso et al., 2002) or malignant melanoma (Musselman et al., 2001) and vaccinations (Kuhlman et al., 2018), and (2) experimental administration of endotoxin in healthy individuals (Eisenberger et al., 2010). Relevant to the current study, a role of striatal dopamine emerged in an IFN-α multi-modal imaging study highlighting decreased dopamine turnover in striatal regions in response to reward, which correlated with depressive symptoms (Capuron et al., 2012).

In addition to structural alterations, inflammation was also shown to impact neural function in MDD. In particular, Dooley et al. (2018) highlighted four endophenotypes of MDD that were affected by experimentally induced inflammation (1) exaggerated reactivity to negative information, (2) increased physiological stress reactivity, (3) increased amygdala responses to social threat, and (4) abnormal reward processing, in particular reduced ventral striatal activation during both anticipation (Eisenberger et al., 2010) and consumption (Capuron et al., 2012; Treadway et al., 2017) of rewards. In a recent functional magnetic resonance imaging (fMRI) study from the sample presented here, we found that stress-induced IL-6 elevations predicted disrupted reward prediction errors in the ventral striatum (Treadway et al., 2017). This endophenotypic approach to examining inflammation emphasizes the importance of understanding its effects on neural structures supporting motivation, affect and cognition, which can be studied in healthy populations to uncover potential mechanisms explaining how inflammation might confer risk for MDD.

The goal of the current study was two-fold. In Study 1, we evaluated whether individual differences in low-grade inflammation [as assessed by baseline IL-6 (as well as CRP)] were associated with volumetric differences in brain regions consistently linked to (1) the effects of inflammation on brain function or structure and (2) phenotypes related to the pathophysiology of MDD, particularly stress regulation and reward/motivation. In Study 2, we leveraged advances in digital medical records and machine learning to derive the presence of MDD in a large clinical dataset, and thereby, test for putative differences in inflammation between patients with v. without probable lifetime MDD and whether inflammation was a predictor of probable MDD classification while accounting for other variables. Based on the literature reviewed above, we hypothesized that increased inflammation in healthy females would be associated with decreased brain volume in the hippocampus, amygdala, ACC, and striatum. Furthermore, we predicted that elevated levels of inflammation from longitudinal medical records would be linked to probable lifetime MDD classification. This work starts with mechanistic data on the effect of inflammation in brain regions previously implicated in depressive phenotypes and reinforces this with medical record data supporting the effect of chronic inflammation on probable lifetime MDD incidence.

Method

Study 1: structural MRI study

Participants

A total of 88 right-handed psychiatrically healthy female participants took part in a larger study that included separate sessions involving a behavioral acute stress manipulation (Admon et al., 2017) and an fMRI study (Treadway et al., 2017). Only females were recruited to avoid potential sex differences in stress response (Kudielka and Kirschbaum, 2005), an important aspect of the overall study (Admon et al., 2017). Participants were excluded for any current or past psychiatric disorder or for five or more lifetime exposures to any recreational drugs (see online Supplementary Methods), as well as any use of any drug or herbal supplement with well-characterized psychotropic effects in the past 3 weeks.

Procedure

Participants were tested in two separate sessions between 11:00 A.M. and 4:00 P.M. to reduce potential circadian effects on endogenous cortisol (Blasovich et al., 2011). At the first session, participants completed an acute laboratory stressor (the Maastricht Acute Stress Test, MAST; Smeets et al., 2012; data reported in Admon et al., 2017). After this first session, participants were asked to return within ~1 month (mean days = 25, S.D. = 21) to complete session 2, which included an fMRI scan (reported in Treadway et al., 2017), as well as a high-resolution anatomical scan.

Plasma collection and IL-6 analysis

To evaluate IL-6 levels in session 1, plasma samples were drawn intravenously three separate timepoints in relation to the acute stressor: >10 min (before stressor), +45 min following stressor and +90 min following stressor (for full details, about the plasma sampling procedure, see Treadway et al., 2017). Only the pre-stress samples were analyzed in order to probe relationships between baseline low-grade inflammation and brain volume (and maximize sample size since all participants with plasma samples provided the first sample).

All assays were performed at the Clinical and Epidemiologic Laboratory (CERLab) within the Department of Laboratory Medicine at Boston Children’s Hospital. IL-6 was assessed using an ultra-sensitive enzyme-linked immunosorbent assay (ELISA; R&D Systems, Minneapolis, MN) employing the quantitative sandwich enzyme immunoassay technique. The assays were run in duplicate, and it was essential that all assays had an inter-assay
coefficient of less than 10%. No IL-6 measurements were excluded due to samples falling outside this predetermined range. The assay had a sensitivity of 0.094 pg/mL, and the average inter-assay coefficient of variation was 7.8%. Based on prior findings (e.g. Treadway et al., 2017; Dooley et al., 2018), IL-6 was our primary inflammatory variable; however, CRP was also assayed to directly link to Study 2.

MRI acquisition and preprocessing
MRI data were acquired at the McLean Imaging Center using a 3T Siemens Tim Trio scanner with a 32-channel head coil. The MRI protocol included high-resolution T1-weighted MPRAGE images (TR = 2200 ms; TE = 1.54 ms; FOV = 230 mm; matrix = 192 × 192; resolution = 1.22 mm³; 144 slices). MRI analysis was carried out using the Computational Anatomy Toolbox (CAT12) (http://www.neuro.unijena.de/cat/) voxel-based morphometry (VBM) module, developed for SPM12 (Wellcome Department of Cognitive Neurology). VBM data were pre-processed as follows: (1) spatial registration to a standard brain template, (2) tissue segmentation into gray and white matter and CSF, (3) bias correction of intensity nonuniformities, and (4) smoothing (8 mm³). For structural MRI, gray matter volumes were extracted from a priori defined regions of interest (ROIs) previously linked to MDD pathophysiology and inflammatory responses: amygdala, hippocampus, and ACC. In addition, exploratory analyses were conducted on volumes of striatal nuclei: caudate nucleus, putamen and NAc. All ROI masks were defined anatomically using the neuromorphometrics atlas (Bakker et al., 2015) as implemented in CAT.

Statistical analysis
For all ROIs, bilateral volume was considered since we had no lat-eral hypotheses. Baseline IL-6 scores were positively skewed, and thus, a log (ln) transformation was used. Potential relations between ROI gray matter volumes and ln-transformed IL-6 levels were examined using Pearson’s correlation in R (R Development Core Team, 2013), separately per region. Spearman’s correlation was also examined to assess monotonic relationship between the variables (on untransformed data). Correction for multiple testing (e.g. Treadway et al., 2017; Dooley et al., 2018), IL-6 was our primary inflammatory variable; however, CRP was also assayed to directly link to Study 2.

MRI acquisition and preprocessing
MRI data were acquired at the McLean Imaging Center using a 3T Siemens Tim Trio scanner with a 32-channel head coil. The MRI protocol included high-resolution T1-weighted MPRAGE images (TR = 2200 ms; TE = 1.54 ms; FOV = 230 mm; matrix = 192 × 192; resolution = 1.22 mm³; 144 slices). MRI analysis was carried out using the Computational Anatomy Toolbox (CAT12) (http://www.neuro.unijena.de/cat/) voxel-based morphometry (VBM) module, developed for SPM12 (Wellcome Department of Cognitive Neurology). VBM data were pre-processed as follows: (1) spatial registration to a standard brain template, (2) tissue segmentation into gray and white matter and CSF, (3) bias correction of intensity nonuniformities, and (4) smoothing (8 mm³). For structural MRI, gray matter volumes were extracted from a priori defined regions of interest (ROIs) previously linked to MDD pathophysiology and inflammatory responses: amygdala, hippocampus, and ACC. In addition, exploratory analyses were conducted on volumes of striatal nuclei: caudate nucleus, putamen and NAc. All ROI masks were defined anatomically using the neuromorphometrics atlas (Bakker et al., 2015) as implemented in CAT.

Statistical analysis
For all ROIs, bilateral volume was considered since we had no lat-eral hypotheses. Baseline IL-6 scores were positively skewed, and thus, a log (ln) transformation was used. Potential relations between ROI gray matter volumes and ln-transformed IL-6 levels were examined using Pearson’s correlation in R (R Development Core Team, 2013), separately per region. Spearman’s correlation was also examined to assess monotonic relationship between the variables (on untransformed data). Correction for multiple comparisons (six ROIs) was applied using the Bonferroni method ($p = 0.05/6 = 0.0083$).

Study 2: biobank study
Participants
Samples and health information were obtained from the Partners HealthCare Biobank, a biorepository of consented patients’ samples at Partners HealthCare (parent organization of Massachusetts General Hospital and Brigham and Women’s Hospital, Boston, USA). The Partners Biobank provides banked samples and laboratory test results collected from patients in Massachusetts who consented to broad-based research. In addition, these test results are linked to clinical data from the Electronic Medical Record (EMR) and survey data on lifestyle, environment, and family history. At time of writing (December 2018), 93,908 patients had provided consent to join the Partners Biobank. De-identified patient data were downloaded using the Biobank Portal, a secure web-based tool that links consented subjects from the Partners Biobank with their healthcare data from the EMR and with research data. Data were extracted from the Partners Biobank on 6 December 2018. Patients were included in the analysis if they had at least one CRP serum test result (see online Supplementary Table S2). Participants were considered to be inflamed if their serum CRP level was above 3 mg/l, in accordance with the accepted norm (Pearson et al., 2003; Bell et al., 2017), and as used in other studies of MDD (Uher et al., 2014; Köhler-Forsberg et al., 2017).

The phenotype of interest in Study 2 categorizes biobank participants into two groups: (1) those with probable lifetime MDD and (2) those without probable lifetime MDD. Curated disease population validated phenotypes defined by bioinformatics algorithms were used to define existence of lifetime MDD at 0.90 positive predictive value. The calculation of these phenotypes is described in Gainer et al. (2016). Briefly, these phenotypes use an analysis file of concepts including potential positive and negative predictors of the disease. These predictors consist of a combination of both coded terms (e.g. prescription of an antidepressant drug) and terms extracted from the narrative data using natural language processing and validation with chart reviews (e.g. the phrase ‘major depression’). An adaptive LASSO penalized logistic regression method identified weighted predictive variables for the algorithm (see online Supplementary Table S3). Most importance was placed on how the variables together in the algorithm could predict the phenotypes, rather than the accuracy of any one variable. Finally, a logistic regression model assigned each biobank participant a probability of having a phenotype based on their values for each term. In development, these predictive values were evaluated in a subsample of a gold-standard patient training set using full chart review by a clinical expert (see online Supplementary Table S4). In addition, computed phenotypes were similarly used to define probable lifetime existence of chronic inflammatory conditions so that patients with these conditions could be excluded from the analysis. Information on body mass index (BMI) and smoking history, which have been each linked to increased inflammation (Bazzano et al., 2003; Nicklas et al., 2004), were also downloaded for inclusion as covariates in regression analyses.

Statistical analysis
The biobank data were analyzed first using non-parametric statistics [Mann–Whitney U test in R (R Development Core Team, 2013)] when the dependent variables (CRP lab count, mean CRP levels) were skewed. Data were also analyzed using logistic regression in R (R Development Core Team, 2013) when the dependent variable was binary (lifetime MDD). Tests were aimed to examine the relationship between probable lifetime MDD and elevated CRP.

Results
Study 1: structural variability and mean IL-6 levels
A total of 53 participants had both IL-6 and structural MRI data. As summarized in Table 1, Pearson’s correlations between ln-transformed baseline IL-6 levels and gray matter volume revealed significant negative correlations in the bilateral hippocampus (Pearson’s $r = −0.38, p = 0.005$), caudate ($r = −0.34, p = 0.01$) and putamen ($r = −0.41, p = 0.002$). These findings remained significant when using non-parametric Spearman’s rank correlations. Thus, increased IL-6 was robustly associated with reduced gray matter in these regions. For the amygdala, the correlation was trending ($r = −0.25, p = 0.07$), whereas no findings emerged for the NAc and the ACC (all $ps > 0.1$). The hippocampus and putamen findings survived Bonferroni
correction. To more directly link to Study 2 analyses were also run with CRP and results were broadly similar to the findings with IL-6 (Fig. 1) (see online Supplementary Table S6).

**Study 2: biobank CRP and curated disease populations**

A total of 12 589 (13.1%) of biobank participants had at least one CRP lab result. A total of 2541 (20.2%) met the criteria for probable current or past MDD (lifetime MDD). In this sample with at least one CRP lab result, at least one chronic inflammatory condition was probable in 8441 (67.1%) participants, consisting of 77.9% who met the criteria for lifetime MDD and in 60.1% of those who did not meet the criteria for probable lifetime MDD (\(\chi^2(1) = 277.88, p < 0.0001\)) (Fig. 2). See online Supplementary Table S5 for frequencies of the nine identified inflammatory conditions.

A \(\chi^2\) test was performed to examine the relationship between probable lifetime MDD and probable lifetime occurrence of the chronic inflammatory medical conditions, yielding a significant effect, \(\chi^2(8) = 114.51, p < 0.001\) \((n = 8441)\). An examination of the residuals and the top three contributions to the overall \(\chi^2\) score suggested that patients with lifetime MDD were more likely to have lifetime asthma (28.3% of total \(\chi^2\) score) and T2 diabetes (17.3% of total \(\chi^2\) score) and less likely to have Crohn’s disease (12.5% of total \(\chi^2\) score), compared to patients without lifetime MDD.

To test the relationship between moderate inflammation and MDD, we first completed analyses for the full sample \([n = 12 589, with 20.2% (n = 2541) with lifetime MDD]\ and then repeated the analysis excluding participants with any of the above chronic inflammatory medical conditions, bringing our sample to \(n = 560\). Next, to examine the differences in lifetime inflammation in the MDD v. no MDD group across all available lab results for a given patient, we considered both mean CRP levels and also the count of all CRP labs that met the cut-off for inflammation (>3 mg/l) for each patient. The data showed a strong positive skew (Fig. 3a, b) and therefore non-parametric statistics were used to compare the groups (lifetime MDD v. no MDD).

For the full dataset a Mann–Whitney–Wilcoxon test indicated that the distributions of the count of elevated CRP labs (>3 mg/l) was also different for participants with \((n = 560)\) v. without \((n = 3588)\) lifetime MDD, \(W = 899 650, p < 0.001\). Again, there were no significant differences in the distributions of mean CRP level across the two groups \((p = 0.20)\).

**Regression analyses**

Smoking history was available for 9129 (72.5%) of participants in the full sample. Lifetime smoking was defined as the inverse of the classification ‘Tobacco use – never’ and includes those participants with tobacco use classified as ‘yes’, ‘passive’, ‘quit’. BMI information was available for the full sample and two categories were classified: those with a BMI ever reaching overweight (>25) or those with a BMI ever reaching underweight (<18.5). Minimum sample size for logistic regression with five predictors and 20.2% proportion of MDD participants was calculated as 247. The full sample was partitioned into training (60%, \(n = 5478\)) and testing (40%, \(n = 3651\)) datasets to allow for subsequent model validation. Logistic regression on the training dataset with a likelihood ratio test revealed that a model including the count of CRP labs >3 mg/l predicted probable MDD classification significantly better than a baseline model including only sex, lifetime smoking and low/high BMI \((\chi^2(6) = 12.86, p < 0.001)\). In this model, for each additional instance of inflammation (CRP lab of >3 mg/l) the predicted odds of being classified with MDD change by a factor of 1.02 (Fig. 4). The addition of mean CRP level as a predictor also significantly improved the model \((\chi^2(7) = 4.33, p = 0.04)\). Examining the absolute value of the \(t\)-statistic for each predictor in the full model (sex + lifetime smoking + low/high BMI + count of elevated CRP labs + mean CRP level), sex was the most important predictor of probable MDD classification \((t = 8.55)\), followed by high BMI \((t = 8.21)\), tobacco use \((t = 6.19)\), low BMI \((t = 5.17)\), count of elevated CRP labs \((t = 3.71)\) and finally mean CRP level \((t = 2.04)\). A Wald \(\chi^2\) determined that inflammation was a significant individual predictor of probable MDD classification \((\chi^2(1,5471) = 13.81, p < 0.001)\) and mean CRP level was also a significant individual predictor of probable MDD classification \((\chi^2(1,5471) = 4.14, p = 0.04)\). Model estimates from the training set were then used to predict values on the testing set. A comparison of the predicted target variable v. the observed values for each observation showed a classification rate of 78.5%.

The logistic regression was then repeated for the reduced dataset \((n = 4148)\). This was again reduced by smoking history being
available for 2809 (67.7%) of participants in the reduced sample. BMI information was available for the full sample. Minimum sample size for logistic regression with the same number of predictors but only 13.5% proportion of MDD participants was calculated as 370. The sample was also partitioned into training (60%, \(n = 1687\)) and testing (40%, \(n = 1122\)) datasets. Logistic regression on the training dataset with a likelihood ratio test revealed that a model including the count of CRP labs \(>3\) mg/l showed a trend to predict probable MDD classification better than a baseline model including only sex, lifetime smoking and low/high BMI (\(\chi^2(6) = 3.55, p = 0.06\)). In this model, for each additional instance of inflammation (CRP lab of \(>3\) mg/l) the predicted odds of being classified with MDD change by a factor of 1.03 (Fig. 4). The addition of mean CRP level as a predictor did not significantly improve the model in this reduced sample (\(\chi^2(7) = 1.49, p = 0.22\)). Examining the absolute value of the \(t\)-statistic for each predictor in the full model (sex + lifetime smoking + low/high BMI + count of elevated CRP labs), sex was the most important predictor of probable MDD classification (\(t = 4.53\)), followed by high BMI (\(t = 3.76\)), tobacco use (\(t = 3.69\)), low BMI (3.54) and finally count of elevated CRP labs (\(t = 2.01\)). A Wald \(\chi^2\) determined that, in this reduced sample, inflammation was still a significant individual predictor of probable MDD classification (\(F_{(1,1681)} = 4.03, p = 0.04\)). Sex did not significantly moderate the effect of inflammation on probable MDD classification (all \(p_s > 0.38\)). Model estimates from the training set were then used to predict values on the testing set. A comparison of the predicted target variable \(v\) the observed values for each observation showed a classification rate of 84.5% in this reduced sample. Thus, although power and significance were diminished in this reduced sample, inflammation was still a noteworthy predictor of probable MDD classification.

**Discussion**

An integration of approaches – including structural imaging and mining of medical records – was used to examine potential mechanisms linked to the role of inflammation in the onset and maintenance of MDD and possible inflammatory laboratory indicators of increased risk for MDD. Here, we observed that increased inflammation was associated with reduced gray matter volume in regions previously linked to endophenotypes...
underlying the pathophysiology of MDD (Treadway and Pizzagalli, 2014), in particular stress regulation (e.g. hippocampus), anhedonia (striatum) and threat reactivity (amygdala). Furthermore, we leveraged a large clinical database to show the predictive relationship between increased lifetime occurrences of elevated inflammation and lifetime incidence of probable MDD, which was established using complex phenotypes emerging from longitudinal medical records using machine-learning approaches. Genome-wide association study data have been used to explore genetic relations between phenotypes for
psychiatric and inflammatory conditions, with an emergent genome-wide correlation between hypothyroidism and MDD (Tylee et al., 2018). However, no study has, to our knowledge, utilized a large database of clinical records and machine-learning computed phenotypes to probe this relationship in an unselected community sample. In Study 1, significant negative correlations emerged between inflammation (measured from serum IL-6) and brain volume in regions linked to MDD endophenotypes and disease course. The hippocampus, in particular, is implicated in the downstream effects of chronic inflammation (Calabrese et al., 2014), as one of the few regions implicated in neurogenesis (Balu and Lucki, 2009; Kempermann et al., 2018), which may be impeded directly by inflammatory cytokines (Koo and Duman, 2008). These findings may provide new insights into how inflammation may trigger depressive symptoms through detrimental effects on neurogenesis or neuroplasticity in regions implicated in stress regulation (e.g. hippocampus), anhedonia (e.g. caudate) and threat reactivity (e.g. amygdala), either directly or through promotion of glucocorticoid secretion after stress (Goshen et al., 2008). This adds to growing literature implicating chronic inflammation in the onset of MDD (Calabrese et al., 2014), as one of the few regions implicated in neurogenesis (Balu and Lucki, 2009; Kempermann et al., 2018), which may be impeded directly by inflammatory cytokines (Koo and Duman, 2008). These findings may provide new insights into how inflammation may trigger depressive symptoms through detrimental effects on neurogenesis or neuroplasticity in regions implicated in stress regulation (e.g. hippocampus), anhedonia (e.g. caudate) and threat reactivity (e.g. amygdala), either directly or through promotion of glucocorticoid secretion after stress (Goshen et al., 2008). This adds to growing literature implicating chronic inflammation in the onset of MDD (Calabrese et al., 2014), as one of the few regions implicated in neurogenesis (Balu and Lucki, 2009; Kempermann et al., 2018), which may be impeded directly by inflammatory cytokines (Koo and Duman, 2008). These findings may provide new insights into how inflammation may trigger depressive symptoms through detrimental effects on neurogenesis or neuroplasticity in regions implicated in stress regulation (e.g. hippocampus), anhedonia (e.g. caudate) and threat reactivity (e.g. amygdala), either directly or through promotion of glucocorticoid secretion after stress (Goshen et al., 2008). This adds to growing literature implicating chronic inflammation in the onset of MDD (Calabrese et al., 2014), as one of the few regions implicated in neurogenesis (Balu and Lucki, 2009; Kempermann et al., 2018), which may be impeded directly by inflammatory cytokines (Koo and Duman, 2008). These findings may provide new insights into how inflammation may trigger depressive symptoms through detrimental effects on neurogenesis or neuroplasticity in regions implicated in stress regulation (e.g. hippocampus), anhedonia (e.g. caudate) and threat reactivity (e.g. amygdala), either directly or through promotion of glucocorticoid secretion after stress (Goshen et al., 2008). This adds to growing literature implicating chronic inflammation in the onset of MDD (Calabrese et al., 2014), as one of the few regions implicated in neurogenesis (Balu and Lucki, 2009; Kempermann et al., 2018), which may be impeded directly by inflammatory cytokines (Koo and Duman, 2008). These findings may provide new insights into how inflammation may trigger depressive symptoms through detrimental effects on neurogenesis or neuroplasticity in regions implicated in stress regulation (e.g. hippocampus), anhedonia (e.g. caudate) and threat reactivity (e.g. amygdala), either directly or through promotion of glucocorticoid secretion after stress (Goshen et al., 2008). This adds to growing literature implicating chronic inflammation in the onset of MDD (Calabrese et al., 2014), as one of the few regions implicated in neurogenesis (Balu and Lucki, 2009; Kempermann et al., 2018), which may be impeded directly by inflammatory cytokines (Koo and Duman, 2008). These findings may provide new insights into how inflammation may trigger depressive symptoms through detrimental effects on neurogenesis or neuroplasticity in regions implicated in stress regulation (e.g. hippocampus), anhedonia (e.g. caudate) and threat reactivity (e.g. amygdala), either directly or through promotion of glucocorticoid secretion after stress (Goshen et al., 2008). This adds to growing literature implicating chronic inflammation in the onset of MDD (Calabrese et al., 2014), as one of the few regions implicated in neurogenesis (Balu and Lucki, 2009; Kempermann et al., 2018), which may be impeded directly by inflammatory cytokines (Koo and Duman, 2008). These findings may provide new insights into how inflammation may trigger depressive symptoms through detrimental effects on neurogenesis or neuroplasticity in regions implicated in stress regulation (e.g. hippocampus), anhedonia (e.g. caudate) and threat reactivity (e.g. amygdala), either directly or through promotion of glucocorticoid secretion after stress (Goshen et al., 2008). This adds to growing literature implicating chronic inflammation in the onset of MDD (Calabrese et al., 2014), as one of the few regions implicated in neurogenesis (Balu and Lucki, 2009; Kempermann et al., 2018), which may be impeded directly by inflammatory cytokines (Koo and Duman, 2008). These findings may provide new insights into how inflammation may trigger depressive symptoms through detrimental effects on neurogenesis or neuroplasticity in regions implicated in stress regulation (e.g. hippocampus), anhedonia (e.g. caudate) and threat reactivity (e.g. amygdala), either directly or through promotion of glucocorticoid secretion after stress (Goshen et al., 2008). This adds to growing literature implicating chronic inflammation in the onset of MDD (Calabrese et al., 2014), as one of the few regions implicated in neurogenesis (Balu and Lucki, 2009; Kempermann et al., 2018), which may be impeded directly by inflammatory cytokines (Koo and Duman, 2008). These findings may provide new insights into how inflammation may trigger depressive symptoms through detrimental effects on neurogenesis or neuroplasticity in regions implicated in stress regulation (e.g. hippocampus), anhedonia (e.g. caudate) and threat reactivity (e.g. amygdala), either directly or through promotion of glucocorticoid secretion after stress (Goshen et al., 2008). This adds to growing literature implicating chronic inflammation in the onset of MDD (Calabrese et al., 2014), as one of the few regions implicated in neurogenesis (Balu and Lucki, 2009; Kempermann et al., 2018), which may be impeded directly by inflammatory cytokines (Koo and Duman, 2008). These findings may provide new insights into how inflammation may trigger depressive symptoms through detrimental effects on neurogenesis or neuroplasticity in regions implicated in stress regulation (e.g. hippocampus), anhedonia (e.g. caudate) and threat reactivity (e.g. amygdala), either directly or through promotion of glucocorticoid secretion after stress (Goshen et al., 2008). This adds to growing literature implicating chronic inflammation in the onset of MDD (Calabrese et al., 2014), as one of the few regions implicated in neurogenesis (Balu and Lucki, 2009; Kempermann et al., 2018), which may be impeded directly by inflammatory cytokines (Koo and Duman, 2008). These findings may provide new insights into how inflammation may trigger depressive symptoms through detrimental effects on neurogenesis or neuroplasticity in regions implicated in stress regulation (e.g. hippocampus), anhedonia (e.g. caudate) and threat reactivity (e.g. amygdala), either directly or through promotion of glucocorticoid secretion after stress (Goshen et al., 2008). This adds to growing literature implicating chronic inflammation in the onset of MDD (Calabrese et al., 2014), as one of the few regions implicated in neurogenesis (Balu and Lucki, 2009; Kempermann et al., 2018), which may be impeded directly by inflammatory cytokines (Koo and Duman, 2008). These findings may provide new insights into how inflammation may trigger depressive symptoms through detrimental effects on neurogenesis or neuroplasticity in regions implicated in stress regulation (e.g. hippocampus), anhedonia (e.g. caudate) and threat reactivity (e.g. amygdala), either directly or through promotion of glucocorticoid secretion after stress (Goshen et al., 2008). This adds to growing literature implicating chronic inflammation in the on...
challenge (e.g. IFN-α treatment). We also acknowledge that our baseline measurements of IL-6 and CRP in Study 1 do not directly show chronic inflammation; moreover, since these measurements were taken in a single session, we cannot comment whether they reflect an acute or chronic inflammatory state. Finally, only females were recruited for Study 1, limiting generalizability. Despite these limitations, convergence between findings from both studies strengthens the association between inflammation and MDD risk. This information could be used to create models predicting the onset of MDD from medical data. More fundamentally, the current integration of findings contributes to a better mechanistic understanding of the hypothesized bidirectional relationship between inflammation and MDD, and ultimately provide clues for new targets for treatment and prevention.

Supplementary material. The supplementary material for this article can be found at https://doi.org/10.1017/S0033291719002940.

Acknowledgements. The authors thank Kerry Ressler for his help with Biobank access, Stefanie Nickels for her advice on statistical analyses and Partners HealthCare Biobank for providing samples and health information data.

Financial support. This project was supported by R01 MH088376 and R37 MH088376 from the National Institute of Mental Health (D.A.P.). The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health. The funding organization had no role in the design and conduct of the study; collection, management, analysis, and interpretation of the data; preparation, review, or approval of the manuscript; and decision to submit the manuscript for publication.

Conflict of interest. Over the past 3 years, D.A.P. has received consulting fees from Akili Interactive Labs, BlackThorn Therapeutics, Boehringer Ingelheim, and Takeda and an honorarium from Alkernes, for activities unrelated to the current paper. No funding from these entities was used to support the current work, and all views expressed are solely those of the authors. All other authors report no biomedical financial interests.

Ethical standards. The authors assert that all procedures contributing to this work comply with the ethical standards of the relevant national and institutional committees on human experimentation and with the Helsinki declaration of 1975, as revised in 2008.

References


